

# Communications to the Editor

## Divergent/Convergent Joint Approach with a Half-Protected Initiator Core To Synthesize Surface-Block Dendrimers

Keigo Aoi, Katsuhito Itoh, and Masahiko Okada\*

Department of Applied Biological Sciences, Faculty of Agricultural Sciences, Nagoya University, Chikusa-ku, Nagoya 464-01, Japan

Received August 19, 1996

Revised Manuscript Received September 8, 1997

This paper describes a novel methodology on macromolecular design of block-type dendrimers having a well-defined structure by a divergent/convergent joint approach. The key to block structures is a new method of postremovable half-protection of an initiator core. A *Sugar Ball*<sup>1</sup> family of amphiphilic AB-type surface-block dendrimers was synthesized by using hemispherical building blocks with protected focal functionality.

Block copolymers are, needless to say, one of the most important categories of macromolecules in a variety of aspects such as polymer synthesis, physical property, and applications as functional materials.<sup>2</sup> Although a number of studies related to dendrimers have appeared,<sup>3</sup> few articles on block-type dendrimers have been reported until now.<sup>3,4</sup> The synthetic procedure of block dendrimers has been almost limited in the convergent method.<sup>5</sup> Considering future development of dendrimer-based smart materials, versatile block dendrimer architecture should have significant meaning. Therefore, in this study, we offer the following methodology to elaborate a surface-block dendrimer. Generally, the method is represented as the sequence (a) hemispherical block construction by a divergent-growth procedure with a half-protected initiator core, (b) dual chemical modification of terminal functional groups, (c) deprotection of center cores, and (d) coupling of two hemispherical blocks at their reactive cores in a convergent way.

This divergent/convergent joint approach was actually performed in order to create *Sugar Ball* derivatives having a surface-block structure. We have recently reported the synthesis of globular artificial glycoconjugate *Sugar Balls* under a new concept of space regulation of sugar residues,<sup>1</sup> which possess molecular information and cell recognition function.<sup>6</sup> The molecular recognition ability of *Sugar Balls* has been successfully demonstrated. In the present AB-type block dendrimer design, one block is used as a cell recognition marker, while the other block will be utilized for additional applications.

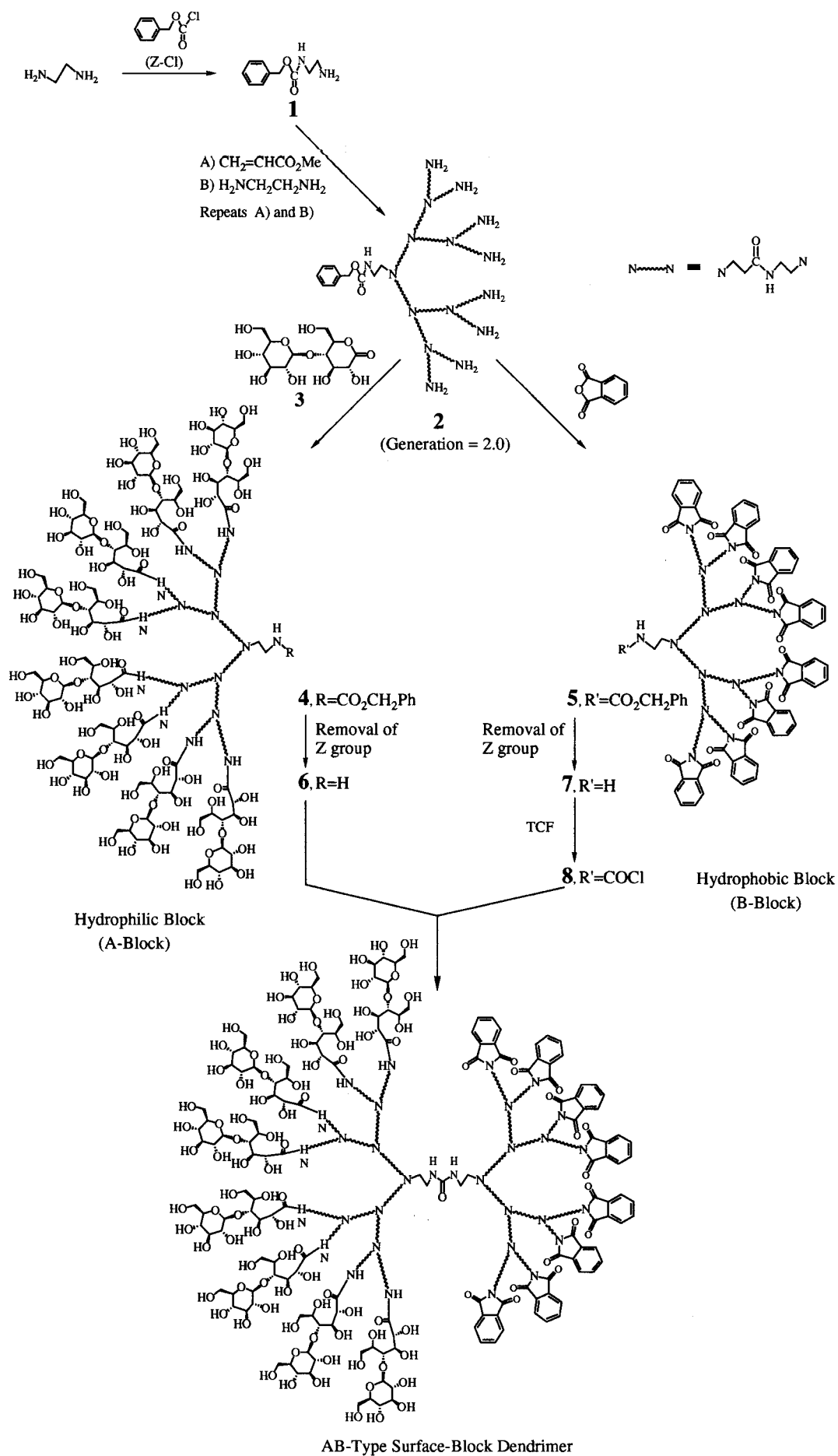
A novel carbohydrate-based AB-type surface-block dendrimer **9** having an amphiphilic structure was synthesized according to Scheme 1. The surface of the A-block is covered with a sugar layer with hydrophilic property. This block is regarded as a model compound

of natural multiantennary oligosaccharides. The surface sector of the B-block provides a hydrophobic part. As a half-protected initiator core, *N*-benzyloxycarbonyl(Z)-protected ethylenediamine **1** was employed.<sup>7</sup> *core*-Z-protected poly(amido amine) (PAMAM) dendrimer (generation 2.0) **2** was prepared by repeated Michael addition and amide formation reactions according to the literature<sup>3a,8</sup> (for a description of *core* and *surface* (*vide infra*), see ref 9). **2** was divided into two portions. One of the advantages of the present method is that both A and B blocks can be derived from a single hemispherical building block. Hydrophilic *surface*-sugar-substituted PAMAM dendrimer **4** was synthesized by the reaction of *surface*-amino groups of **2** with a 38-fold molar excess *O*- $\alpha$ -D-glucopyranosyl-(1 $\rightarrow$ 4)-D-glucono-1,5-lactone (maltono lactone, **3**) in dimethyl sulfoxide at 45 °C for 20 h under a nitrogen atmosphere.<sup>10</sup> Hydrophobic *surface*-phthaloyl block **5** was derived from **2** and phthalic anhydride (50-fold molar excess, in dimethyl sulfoxide at 100 °C for 30 min. Yield, 98%).<sup>11</sup> *core*-Z-protecting groups of **4** and **5** were selectively removed by hydrogenolysis to produce dendritic fragments with a functional core, **6** and **7**, respectively.<sup>12</sup> *core*-Amino hydrophobic block **7** was treated with trichloromethyl chloroformate (TCF) to give **8** having a reactive core of acid chloride.<sup>13</sup> The equimolar reaction between *core*-amino block **6** and *core*-acid chloride-type **8** was undertaken in dimethyl sulfoxide at 45 °C under a dry condition. The progress of the reaction was followed by <sup>1</sup>H NMR spectroscopy. AB-type surface-block dendrimer **9** was obtained in 45% yield after purification by preparative size exclusion chromatography (SEC) in dimethyl sulfoxide and a subsequent reprecipitation procedure.<sup>14</sup>

In order to check molecular recognition potential of *surface*-sugar-substituted dendritic fragments, the interaction between **4** and concanavalin A lectin was examined by a UV/vis spectrophotometer.<sup>1a,15</sup> The sugar-bearing hemispherical block of generation 2.0 showed apparent recognition ability toward the protein receptor. This fact suggests that polymers with hemispherical blocks derived from **4** have a capability as cell-recognizable biomedical materials.

Dendritic blocks **6–8** are important as building synthons carrying a reactive center core in various dendrimer-based macromolecular designs such as segment-block dendrimers<sup>4a</sup> and block copolymers between dendrimers and linear polymers.<sup>16</sup> In other words, the half-protected initiator core method should be essential in research and development of sophisticated dendritic functional materials, e.g., adhesives, surfactants, and drug delivery systems. For example, **1** will be applicable to a polyamine dendrimer that has been used for a Dendritic Box.<sup>17</sup> Including further extension of the partly protected initiator system, the present architectural principle of AB-type surface-block dendrimer provides numerous applications such as supramolecular self-assembling globular amphiphiles and intelligent nanocapsules having dual binding surface sectors to proteins and DNAs.

Scheme 1



**Acknowledgment.** Financial support from the Ministry of Education, Science, and Culture of Japan (Grant-in-Aid nos. 08246106, 08231232, 08875184, and 09238217) is gratefully acknowledged.

## References and Notes

- (1) (a) Aoi, K.; Itoh, K.; Okada, M. *Macromolecules* **1995**, *28*, 5391. (b) Aoi, K.; Tsutsumiuchi, K.; Yamamoto, A.; Okada, M. *Macromol. Rapid Commun.*, in press. (c) Aoi, K.; Tsutsumiuchi, K.; Yamamoto, A.; Okada, M. *Tetrahedron*, in press.
- (2) (a) Ishizu, K. In *Polymeric Materials Encyclopedia*; Salamone, J. C., Ed.; CRC Press: Boca Raton, FL, 1996; p 783. (b) Yagci Y.; Mishra, M. K. *Ibid.*, p 789. (c) Faradet, A. *Ibid.*, p 797.
- (3) (a) Tomalia, D. A.; Dvornic, P. R. In *Polymeric Materials Encyclopedia*; Salamone, J. C., Ed.; CRC Press: Boca Raton, FL, 1996; p 1814. (b) Fréchet, J. M. J.; Hawker, C. J. In *Comprehensive Polymer Science*, second supplement, Allen, G., Ed.; Pergamon, Elsevier Science: Oxford, U.K., 1996, p 71. (c) Newkome, G. R.; Moorefield, C. N.; Vögtle, F. *Dendritic Molecules: Concepts, Synthesis, Perspectives*; VCH: Weinheim, 1996.
- (4) (a) Hawker, C. J.; Wooley, K. L.; Fréchet, J. M. J. *Macromol. Symp.* **1994**, *77*, 11. (b) Hawker, C. J.; Fréchet, J. M. J. *Macromolecules* **1990**, *23*, 4726. (c) Wooley, K. L.; Hawker, C. J.; Fréchet, J. M. J. *J. Chem. Soc., Perkin Trans. 1* **1991**, 1059. (d) Hawker, C. J.; Fréchet, J. M. J. *J. Am. Chem. Soc.* **1992**, *114*, 8405.
- (5) Tomalia, D. A.; Swanson, D. R.; Klimash, J. W.; Brothers, H. M., III. *Polym. Prepr. (Am. Chem. Soc., Div. Polym. Chem.)* **1993**, *34*, 52.
- (6) (a) Aoi, K.; Suzuki, H.; Okada, M. *Macromolecules* **1992**, *25*, 7073. (b) Aoi, K.; Tsutsumiuchi, K.; Okada, M. *Macromolecules* **1994**, *27*, 875. (c) Aoi, K.; Tsutsumiuchi, K.; Aoki, E.; Okada, M. *Macromolecules* **1996**, *29*, 4456.
- (7) **1** was prepared by the reaction of benzyloxycarbonyl chloride with a 20-fold molar excess of ethylenediamine in methanol with mixing at  $-50^{\circ}\text{C}$ , then raising to  $-5^{\circ}\text{C}$  for 20 h (yield, 66%).
- (8) Michael addition between amino compounds, e.g., **1**, and methyl acrylate was carried out in methanol at  $40^{\circ}\text{C}$  for 120 h. The resulting methyl ester derivatives were allowed to react with ethylenediamine in methanol at  $27^{\circ}\text{C}$  for 72 h. Each half-generational product was purified by preparative flash-column chromatography using an acetone eluent with addition of methanol gradationally.
- (9) By the conventional nomenclature, positions at an initiating end and a terminal end of the linear polymer are denoted as  $\alpha$  and  $\omega$ , respectively. In this nomenclature, positions of a center core and terminal branches are determined with respect to divergent and convergent methods in dendrimer synthesis, that is,  $\alpha$  and  $\omega$  for the former method and  $\omega$  and  $\alpha$  for the latter method, respectively. Thus, in dendrimer chemistry, the authors propose terms *core*- and *surface*-, which indicate the positions of a center core and terminal branches straightforwardly. While the term of reactive focal point is well-known, this does not cover the nonreactive center core of globular dendrimer.
- (10) **4** was purified by dialysis against water using a cellulose tube (Spectrum Medical Industries, Inc., MW cutoff 1000) (yield, 34%).
- (11) *core*-Benzyloxycarbonyl-*surface*-perphthaloylpoly(amido amine) dendrimer **5** ( $G = 2.0$ ): IR (KBr) 3100 ( $\nu_{\text{C-H}}$ ), 2950 ( $\nu_{\text{C-H}}$ ), 1780, 1720 ( $\nu_{\text{C=O}}$ , imide), 1640 ( $\nu_{\text{C=O}}$ , amide), 1560 ( $\delta_{\text{N-H}}$ , amide), 750, 720 ( $\delta_{\text{C-H}}$ , amide)  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR ( $(\text{CD}_3)_2\text{SO}$ , TMS,  $27^{\circ}\text{C}$ , 270 MHz)  $\delta$  8.10 (br, NH), 7.86–7.53 (m, CH of phthalimide), 7.33 (m, aromatic CH of Z), 5.02 (s,  $\text{CH}_2$  of Z), 3.64 (m,  $\text{CH}_2$  adjacent to phthalimide nitrogen), 3.26 (m,  $\text{NHCH}_2$  of PAMAM), 3.05 (m,  $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 2.45 (m,  $\text{NCH}_2\text{CH}_2\text{CONH}$  and  $\text{CH}_2\text{CONH}$  of PAMAM);  $^{13}\text{C}$  NMR ( $(\text{CD}_3)_2\text{SO}$ , TMS,  $27^{\circ}\text{C}$ , 100 MHz)  $\delta$  170.8 (C=O, amide), 168.4 (C=O, phthalimide), 156.3 (C=O, Z), 138.4 (quaternary aromatic carbon of Z), 134.2–122.8 (aromatic carbon), 65.4 ( $\text{CH}_2$  of Z), 51.2 ( $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 49.1 ( $\text{NCH}_2\text{CH}_2\text{CONH}$  of PAMAM), 37.3 ( $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 30.9 ( $\text{CH}_2\text{CONH}$  of PAMAM).
- (12) **4** was treated with hydrogen in the presence of a Pd/C catalyst in a 50/50 mixture of methanol and water at  $27^{\circ}\text{C}$  to give **6** (yield, 72%). Similar deprotection of the Z group of **5** was conducted with  $\text{H}_2/\text{Pd}-\text{C}$  in methanol at  $27^{\circ}\text{C}$  (7, 84% yield). Removal of the Z group was also conducted by using a trifluoroacetic acid solution containing 7 wt % of thioanisole at  $27^{\circ}\text{C}$ .
- (13) Reaction conditions: in chloroform at  $45^{\circ}\text{C}$  under nitrogen. **8** was purified by repeated reprecipitations (chloroform (solvent)/diethyl ether (nonsolvent) under dry conditions.
- (14) *surface-block*-Permaltoibionamido/perphthaloylpoly(amido amine) dendrimer **9** ( $G = 2.0$ ):  $M_w/M_n$  (SEC,  $27^{\circ}\text{C}$ ,  $\text{Me}_2\text{SO}$ ), 1.01; IR (KBr) 3650–3100 ( $\nu_{\text{N-H}}$  and  $\nu_{\text{O-H}}$ ), 3100 ( $\nu_{\text{C-H}}$ ), 2950 ( $\nu_{\text{C-H}}$ ), 1780, 1720 ( $\nu_{\text{C=O}}$ , imide), 1645 ( $\nu_{\text{C=O}}$ , amide), 1635 ( $\nu_{\text{C=O}}$ , urea), 1550 ( $\delta_{\text{N-H}}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR ( $(\text{CD}_3)_2\text{SO}$ , TMS,  $27^{\circ}\text{C}$ , 400 MHz)  $\delta$  8.10, 8.04, 8.01 (br, NH), 7.91–7.69 (m, CH of phthalimide), 4.95–3.32 (m, sugar residue), 3.23 (m,  $\text{NHCH}_2$  of PAMAM), 3.00 (m,  $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 2.41 (m,  $\text{NCH}_2\text{CH}_2\text{CONH}$  and  $\text{CH}_2\text{CONH}$  of PAMAM);  $^{13}\text{C}$  NMR ( $(\text{CD}_3)_2\text{SO}$ , TMS,  $27^{\circ}\text{C}$ , 100 MHz)  $\delta$  174.1 (C=O of maltobionamide), 170.9 (C=O, amide), 168.2 (C=O, phthalimide), 153.2 (C=O, urea), 134.2–122.9 (aromatic carbon), 100.6 (anomeric carbon), 73.2, 73.1, 72.2, 72.0, 71.9, 71.5, 70.7, 69.8 (other carbons derived from maltose lactone), 51.8 ( $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 48.5 ( $\text{NCH}_2\text{CH}_2\text{CONH}$  of PAMAM), 38.1 ( $\text{NHCH}_2\text{CH}_2\text{N}$  of PAMAM), 31.3 ( $\text{CH}_2\text{CONH}$  of PAMAM).
- (15) Precipitation studies were carried out by mixing equivolume phosphate buffer solutions (pH 7.4) of concanavalin A (Con A, Sigma Chemical Co., 6 mg/mL) and **4**. Turbidity was estimated by a UV/vis spectrophotometer. Solutions of **4** (2.0, 1.0, and 0.5 mg/mL) were added to the Con A solutions, and absorbance values were 0.01, 0.21, and 0.44, respectively, after incubation at  $25^{\circ}\text{C}$  for 20 min. Precipitation is caused by the formation of cross-linking between Con A and glucose residues of antennary synthetic glycoconjugate **4**.
- (16) (a) Aoi, K.; Motoda, A.; Okada, M.; Imae, T. *Macromol. Rapid Commun.* **1997**, *18*, 945. (b) Leduc, M. R.; Hawker, C. J.; Dao, J.; Fréchet, J. M. J. *J. Am. Chem. Soc.* **1996**, *118*, 11111. (c) van Hest, J. C. M.; Baars, M. W. P. L.; Eissen-Roman, C.; van Genderen, M. H. P.; Meijer, E. W. *Macromolecules* **1995**, *28*, 6689. (d) Chapman, T. M.; Hillyer, G. L.; Mahan, E. J.; Shaffer, K. A. *J. Am. Chem. Soc.* **1994**, *116*, 11195.
- (17) Jansen, J. F. G. A.; de Brabander-van den Berg, E. M. M.; Meijer, E. W. *Science* **1994**, *266*, 1226.

MA961397N